**Review Article** 

Trends in Pharmaceutical Sciences 2019: 5(1): 7-24. Hydrogels as Drug Delivery Systems; Pros and Cons

Parisa Ghasemiyeh<sup>1,2</sup>, Soliman Mohammadi-Samani<sup>2,3</sup>

<sup>1</sup>Department of Clinical Pharmacy, School of Pharmacy, Shiraz University of Medical Sciences, Shiraz, Iran.

<sup>2</sup>Department of Pharmaceutics, School of Pharmacy, Shiraz University of Medical Sciences, Shiraz, Iran.

<sup>3</sup>Center for Nanotechnology in Drug delivery, School of Pharmacy, Shiraz University of Medical Sciences, Shiraz, Iran.

# Abstract

Hydrogels are cross-linked polymers with hydrophilic groups which enable them to absorb large amounts of water. Although hydrogels have numerous capability and advantages in drug delivery including biocompatibility, low toxicity and good swelling behavior but depending on chemical moieties of the gel forming polymers and route of administration some limitations would appear in delivery of active pharmaceutical using hydrogel as delivery vehicle. In this review at first classification of the hydrogel with different approaches including chemical moieties, crosslinking agent behaviors and release controller mechanism was performed and limitations arise from each category was described and finally different approaches to overcome each of this limitation was proposed.

Keywords: Drug delivery, Hydrogels, Polymers, Pros and Cons.

### **1. Introduction**

Hydrogels are novel drug delivery systems which have been considered since the early 1960s. First of all Wichterle and Lim introduced a kind of hydrophobic gel, cross-linked hydroxyethyl methacrylate (HEMA) hydrogels, developed for biological purposes (1, 2). Hydrogels have many advantages such as their biocompatibility potential, hydrophilicity, controlled drug release and smart drug delivery, etc. (3). So, there was a great interest among scientists of different fields to develop and progress these delivery systems. Hydrogels are hydrophilic polymers with cross-links which form a polymeric network enable them to absorb water from 10-20 percent up to thousands times of their own weight (2, 4). There are two kinds of hydrogels; chemically cross-linked or permanent hydrogels, which are stable to degradation during

*Corresponding Author*: Soliman Mohammadi-Samani, Department of Pharmaceutics, School of Pharmacy, Shiraz University of Medical Sciences, Shiraz, Iran. Email: smsamani@sums.ac.ir swelling, and physically cross-linked or reversible hydrogels, which degrade and dissolve during water absorption (1, 2). In chemically cross-linked hydrogels, polymers are cross-linked with covalent bonds (5), but in physically cross-linked hydrogels, polymers are bond together with hydrogen bonds, ionic bonds or hydrophobic bonds (6). The ability of water absorption and swelling of hydrogels results in the presence of hydrophilic moieties in their polymeric structure but their resistance in water dissolution is the outcome of the presence of cross-linkers between the polymeric chains (7). Polymers which are used in hydrogel preparation would be from natural or synthetic sources, each of these types of polymers could have pros and cons and they should be selected according to hydrogels application and target site of drug delivery.

### 2. Hydrogels classification

Hydrogels may be classified based on their sources (natural or synthetic hydrogel), their

polymeric compositions (homo-polymer, co-polymer and multi-polymer hydrogels), physical structure (amorphous, semi-crystalline and crystalline hydrogels), cross-linkers (physical and chemical cross-linked hydrogels), electrical charge (nonionic, cationic, anionic, amphoteric and zwitterionic hydrogels) and release controllers (timecontrolled hydrogels and stimuli-induced or smart hydrogels) (1, 7). (Figure 1)

### 3. Release mechanisms

### 3.1. Diffusion-controlled

Diffusion-controlled release is the most common mechanism of drug release from hydrogels. In this type of drug release, Fick's diffusion theory is used for kinetic modeling (8-11). For porous hydrogels with pore sizes of larger than drug molecule dimensions, drug diffusion from hydrogels could be related to the porosity and also the tortuosity of hydrogels. Diffusion-controlled release hydrogels might be act as reservoir or matrix. In reservoir drug delivery systems, drug molecules are encapsulated and surrounded by polymeric hydrogels and so drug release mostly obeys the first law of Fickian diffusion (8). In matrix drug delivery systems, drug molecules are homogenously dispersed in polymeric hydrogels and drug release mostly follows the second law of Fickian diffusion.

### 3.2. Swelling-controlled

Swelling-controlled drug release could occur when the rate of dug diffusion is faster than the rate of hydrogel swelling. For purely swellingcontrolled drug release, the kinetic model of release could be mostly fit to zero-order model (8, 11). Hydrogels may have swelling-induced transition phase (at glass transition temperature or Tg) from glassy to rubbery state which causes faster drug diffusion and release from polymeric chains. In swelling-controlled delivery systems, the higher the rate of hydrogel swelling, the higher the rate of drug release, so the rate and ability of hydrogels' water absorption and the thickness of polymeric gels are important factors in swelling-controlled delivery systems (8, 12, 13).

### 3.3. Chemically-controlled

Chemically-controlled drug release mechanism explains the mechanism in which a reaction occurs within the hydrogel matrix. In these reactions enzymatic or hydrolytic cleavage of polymeric network is responsible for drug release. Drug release in chemically-controlled delivery systems could be occured by cleavage of polymeric chains through bulk or surface erosion and following these mechanisms, the entrapped drug or tethered drug would be released from hydrogels (8, 10, 11, 14). The polymer chain cleavage is the rate-limiting step of chemically-controlled



### Figure 1. Hydrogels classification.

8

Hydrogels Pros and Cons

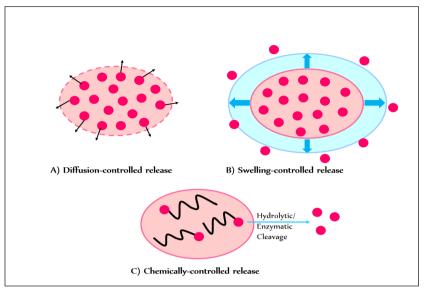


Figure 2. Schematic view of drug release mechanisms from hydrogels; A) diffusion-controlled release, B) swelling-controlled release, and C) chemically-controlled release.

release systems and in this system, drug molecular diffusion has no significant effect on drug release (Figure 2).

## 4. Structure of hydrogels

Hydrogels could be prepared from natural or synthetic polymers. The monomers which are used in hydrogel preparation would be hydrophilic or hydrophobic. Generally, any method used to cross-link the polymers, could be used in hydrogel preparation. Polymers might be cross-linked through chemical or physical reactions or through ionizing radiation method (7). The most important ingredients in hydrogel preparation are monomers, cross-linkers and initiators. According to types of preparation, hydrogels are categorized as the following (Figure 3):

### 4.1. Homo-polymeric network

Homo-polymeric networks are formed from a single type of monomers. According to the nature of monomers and the polymerization technique, it would be possible for homo-polymers to have cross-linked structures (7).

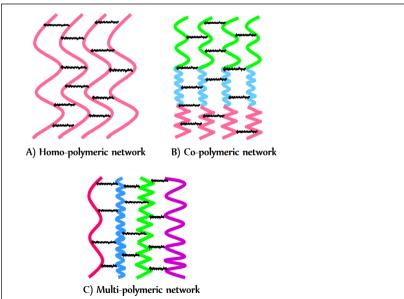


Figure 3. Schematic view of hydrogels categories according to methods of preparation; A) homo-polymeric network hydrogels, B) co-polymeric network, and C) multi-polymeric network hydrogels.

### 4.2. Co-polymeric network

Co-polymeric hydrogels are formed from two or more than two types of monomers. In these hydrogels, the presence of one or more hydrophilic component is necessary (7). The hydrophilic moiety in copolymers is necessary to obtain the swelling ability of hydrogels (15).

### 4.3. Multi-polymeric network

Multi-polymeric hydrogels are formed from two independent cross-linked polymers (7). These dual polymers have amazing properties superior than each of the single polymers. Multipolymeric hydrogels are developed to interpenetrating (IPN) hydrogels which are the combination of two polymers in a network and at least one of these polymers is cross-linked or synthesized (15).

## **5. Polymers used in hydrogel preparation** 5.1. Natural polymers

Natural polymers usually have the advantage of biocompatibility and biodegradability, but because of the distress of purification, their most common limitation is batch to batch variation which causes differences in final formulation (16). Some of the most important natural polymers which are used in hydrogel preparation are listed in the following section.

### 5.1.1. Chitosan

Chitosan is a well-known natural polymer and has many advantages in drug delivery systems such as mucoadhesive and bioadhesive properties, absorption-enhancing properties and controlled drug release (16). Chitosan is a water soluble (in light acidic pH) and cationic (positive charge) polymer. The most important advantages of this polymer are its biocompatibility and low toxicity potential (1). Chitosan also has some limitations such as small specific surface area and void fraction that should be overcome (17).

### 5.1.2. Hyaluronic acid

Hyaluronic acid is a mucopolysacharide which is naturally existing in cartilage and connective tissue. Hyaluronic acid has a poly-anionic nature and so it could be cross-linked to cationic polymers. Hyaluronic acid could be degraded by hyaluronidase (18). Hyaluronic acid is a suitable choice in local delivery because of its biocompatible and biodegradable nature and visco-elastic and unique rheological properties. It could induce cell mobility and cell proliferation and also cause wound healing. Hyaluronic acid is used in hydrogel preparation and has the advantage of delayed drug release and prolonged duration of action (19).

### 5.1.3. Carrageenan

Carrageenan is a natural polysaccharide and because of its gelation capability has the advantage of controlled drug release. Carrageenan is an anionic, biocompatible and low toxic polymer. It seems that  $\kappa$ -carrageenan is the most suitable form of carrageenan for drug delivery and tissue engineering purposes (20).

## 5.1.4. Alginic acid

Alginic acid is an anionic polysaccharide which is highly water soluble and has the gelation capability. It has the advantage of biocompatibility and low toxicity (1). Alginic acid could instantly form a gel in combination with calcium ions (21) and could be used as drug carrier.

# 5.1.5. Collagen

Collagen is a biocompatible and biodegradable polymer which is widely used in pharmaceutical industries (22). It is a very suitable choice for implant preparation. Collagen could control drug release from hydrogels and also could induce cell growth (23). Collagen would be widely used as a scaffold for tissue engineering purposes.

# 5.2. Synthetic polymers

### 5.2.1. Poly ethylene glycol (PEG)

Polyethylene glycol is a synthetic and bio-functional polymer. PEG has the advantage of biocompatibility and adaptable physical properties. PEG is capable to form controlled gelation responsive to a photo-initiator or by mixing it with cross-linkers (24). PEG hydrogels have degradable (hydrolysable) moieties in their structures.

### 5.2.2. Poly lactic acid (PLA)

Poly lactic acid is a synthetic, biodegrad-

Hydrogels Pros and Cons

able, biocompatible and hydrophobic polymer. PLA has the limitation of dispersing hydrophilic materials in their polymeric structures and uncontrolled drug release pattern. These problems could be solved by cross-linking PLA to hydrophilic PEG polymers which is strongly hydrated and capable to sustain and prolong drug release. PEG/ PLA copolymerization has the advantage of both hydrophilic and biodegradable polymers. PEG grafting to PLA induces hydrophilicity in the polymer and PLA fraction of the molecule induces the biodegradability on it (25).

# 5.2.3. Poly lactic co-glycolic acid (PLGA)

PLGA is a biocompatible, biodegradable polymer with suitable mechanical characteristics. PLGA could be used in drug delivery systems because of the advantage of sustained and prolonged drug release (26). Although PLGA is a biodegradable polymer, but micro environmental acidity following polymer degradation, could induce irritation at the site of formulation application and could also damage peptide/protein drugs which are loaded to hydrogels (16).

### 5.2.4. Poly vinyl alcohol (PVA)

PVA is a biodegradable, biocompatible, hydrophilic and synthetic polymer. PVA would be widely used in tissue engineering to repair damaged organs or tissues. Using the optimal PVA to water ratio could produce a suitable hydrogel which could simulate natural tissues properties (27).

### 5.2.5. Poly caprolactone (PCL)

PCL is a biocompatible, biodegradable and hydrophobic synthetic polyester which is highly used in drug delivery and tissue engineering (28). PCL has the advantage of adoptability to physical, mechanical and chemical changes because of the ease and potential of its co-polymerization with other polymers. PCL is compatible with both natural and synthetic polymers. Because of the hydrophobic nature, lipophilic drugs could be homogenously dispersed in polymeric matrix, but hydrophilic drugs mostly migrate to the surface of PCL polymers (29).

# 5.3. Semisynthetic polymers (Cellulose derivatives)

# 5.3.1. Carboxymethyl cellulose (CMC)

CMC is a biocompatible and biodegradable polymer which is widely used for biomedical purposes. CMC is a water-soluble derivative of cellulose (ether of cellulose) and is now available in market with low cost and high purity (30). CMC could be easily cross-linked to produce hydrogels.

# 5.3.2. Hydroxyethyl cellulose (HEC)

HEC is a water soluble and biocompatible derivative of cellulose. HEC has many OH groups in its structure and has the advantage of being modified and improved by grafting polymerization with vinyl groups (31). HEC has better swelling properties when cross-linked with CMC and forms novel hydrogels. The water absorption and swelling ability of these hydrogels could be optimized by adjusting suitable HEC/CMC ratio and amount of cross-linkers used in hydrogel preparation (32).

### 5.3.3. Methyl cellulose (MC)

MC is a semi-flexible linear derivative of cellulose and some of the hydroxyl groups are substituted with methoxy groups. The ratio of hydroxyl to methoxy groups could prescribe the water solubility of this polymer. The gelation of MC could be occurred at sol-gel transition temperature (Tg) by nucleation and growth mechanism (33).

### 5.3.4. Hydroxypropyl cellulose (HPC)

HPC is a highly water soluble derivative of cellulose. HPC hydrogels are usually used in dye removal of aqueous solutions. The most important limitation of HPC hydrogels is their low capacity of water absorption which could be overcome by incorporation of nano-fillers with large specific surface area within these hydrogels (34).

# 5.3.5. Hydroxypropyl methyl cellulose (HPMC)

HPMC is a water soluble derivative of cellulose which is widely used in controlled drug delivery systems. The most important characteristic of HPMC is the high swelling capability that could control the release of incorporated active pharmaceutical ingredient. Physicochemical properties of HPMC polymers could be described according to

the molecular weight, methoxy substitute content and hydroxypropoxy substitute content in their structures (13).

# 6. Hydrogel applications and different routes of administration

Hydrogels could be used as drug delivery systems through different routes of administration as listed in Table 1.

# 6.1. Oral route

Oral drug delivery is the most common

and desirable route of drug administration and has the best patient compliance. Co-polymer hydrogel networks act as a suitable carrier for oral drug delivery, since they can improve oral absorption and bioavailability. Hydrogels are considered as safe drug delivery system for oral route of administration and also have the muco-adhesive capability which could prolong drug release and absorption. The other advantage of hydrogels as oral drug delivery systems is the ability of the protection of incorporated drug from enzymatic degradation (35). Hydrogels are mostly studied in oral drug delivery

Table 1. Different loaded drugs and routes of administration of hydrogels.

	Route of Adminis- tration	Loaded Drug	First Author	The major goal of the study	Ref.
1	Oral	-	Paolo Colombo	Study the use of hydrogel matrices or swelling-controlled de- livery systems for oral drug delivery.	(46)
		-	Kim Knuth	Study the use of hydrogels for drug delivery to the vaginal and oral areas (oral cavity, stomach, small, intestine, colon and rectum).	(47)
		Flavin mono- nucleotide	Waleed S.W. Shalaby	Using enzyme-digestible hydrogels for once daily oral drug delivery by long-term gastric retention potential.	(48)
		Insulin	Mariko Morishita	Using cross-linked polymer microparticles for oral insulin delivery.	(35)
		Insulin	Kiran Chaturvedi	Study the use of hydrogel-based devices for oral insulin de- livery.	(36)
		Insulin	Bumsang Kim	Using photopolymerized pH-responsive hydrogels for oral insulin delivery with the potential of protective effect and change in insulin release rate.	(49)
		Chlorhexidine gluconate	S. S, enel	Using chlorhexidine incorporated in chitosan hydrogel for oral candidiasis.	(50)
2	Paren- teral	Tyramine	K.S. Kim	Using injectable radical cross-linked hyaluronic acid-tyra- mine hydrogels in rheumatoid arthritis.	(51)
		rTIMP-3	Brendan P. Purcell	Using bio-responsive injectable hydrogels for the purpose of as-needed matrix metalloproteinase inhibition to prevent dif- ferent pathologies.	(52)
		-	Amit Alexander	Study the use of thermosensitive injectable hydrogels for biomedical applications.	(37)
		Tramadol	Lorina Bisharat	Study the release of tramadol from Poloxamer thermo-sen- sitive hydrogels.	(53)
		Insulin	Sabrine S. Jensen	Evaluating the release of insulin implants in a hydrogel ma- trix.	(54)
		Peptide and protein	Mayura Oak	Study the use of "smart hydrogels" for controlled parenteral peptide and protein delivery.	(55)
		Protein	Tina Vermonden	Study the use of hydrogels for protein delivery.	(56)

	Route of Adminis- tration	Loaded Drug	First Author	The major goal of the study	Ref.
3	Nasal	-	Jie Wu	Using PEG and chitosan-based thermosensitive hydrogels for nasal drug delivery.	(57)
		-	H. Nazar	Using N-trimethyl chitosan-based thermosensitive hydrogel for nasal drug delivery.	(38)
		-	Barbara Luppi	Study the use of chitosan-based hydrogels for nasal drug de- livery.	(58)
		-	Xiaofeng Xu	Using poloxamer and $\mathcal{E}$ -poly lysine-based thermosensitive and mucoadhesive hydrogels for nasal drug delivery.	(59)
		Phenylephrine hydrochloride	Ibrahim A. Alsarra	Using poly(vinylpyrrolidone) hydrogels for nasal mucosal drug delivery.	(60)
		Acyclovir	Ibrahim A. Alsarra	Using mucoadhesive poly(vinylpyrrolidone) hydrogels for nasal delivery of acyclovir.	(61)
		Antipsychotic drug	Barbara Luppi	Using chitosan/pectin based hydrogel nasal inserts to im- prove the bioavailability of antipsychotic agents.	(62)
4	Ocular	-	M. Zignani	Study the use of semi-solid ophthalmic viscose and mucoad- hesive hydrogels to improve ocular residence time and drug bioavailability.	(63)
		-	Derya Gulsen	Preparation of a dispersion of microemulsions drops in poly- 2-hydroxyethyl methacrylate (p-HEMA) hydrogels for the purpose of ocular drug delivery.	(39)
		Acetazol- amide	Andreza Ribeiro	Using HEMA polymer as a backbone component with the potential of cytocompatible and biomimetic properties for controlled ocular drug delivery.	(64)
		-	Li Xinming	Study the use of polymeric hydrogels for ocular drug deliv- ery through contact lens.	(65)
		-	Eugen Barbu	Preparation of nanoparticulate hybrid polymeric hydrogels with both thermosensitive and swelling properties for ocular drug delivery.	
		-	Xiaohong Hu	Study the use of hydrogel contact lens for the purpose of con- trolled ocular drug delivery.	
		-	HongBo Yin	Synthesis of a biodegradable and thermosensitive poly(ethylene glycol)-poly(e-caprolactone)-poly-(ethylene glycol) (PEG-PCL-PEG) (PECE) triblock copolymer in situ hydrogel for sustained ocular drug delivery.	(67)
5	Topical	Betametha- sone-17-val- erate	Taner S, enyi git	Using topical deoxycholate hydrogels of betamethasone- 17-valerate in order to avoid skin irritation.	
		-	Claudia Valenta	Study the use of Deoxycholate-hydrogels as novel topical drug delivery systems.	

Route of Administration	of Loaded 5- Drug	First Author	The major goal of the study	Re
	Propranolol hydrochloride	T. Cerchiara	Formation of physically cross-linked chitosan hydrogels with lauric, myristic, palmitic or stearic acid for topical de- livery of hydrophilic agents.	(7(
	Silver sulfa- diazine Econazole	Nadia M. Morsi Vanna Sanna	Using Silver sulfadiazine based cubosome hydrogels (cubo- gels) in order to treat deep second degree burns. Preparation of solid lipid nanoparticles incorporated to	
	nitrate Nonivamide	Jia-You Fang	HPMC hydrogels for topical delivery of econazole nitrate. Using chitosan and carboxymethylcellulose hydrogels of nonivamide to improve skin permeation and distribution.	
	Isosorbide mononitrate	Yoncheva, Krassimira Diarmaid J. Murphy	Using Carbopol 940-based hydrogels for topical delivery of isosorbide mononitrate for the treatment of fissures. Evaluation of the efficay of poly(vinyl alcohol) tetrahydroxy-	(7 (7
	Terbinafine hydrochloride	İpek Özcan	borate hydrogels as topical drug delivery systems.	(7
Brain de livery	e- Peptide or aminosugar	Giles W. Plant	Evaluation of the use of synthetic hydrogels containing the sequence of arginine–glycine–aspartic acid (RGD) peptide and aminosugars, which are found in many Extracellular matrix glycoproteins.	(7
••••••	-	K. B. Bjugstad	Using PEG-based hydrogels implants to provide tissue engi- neering and sustained drug release in central nervous system (localized recruitment and activation of microglia and astro- cytes).	(7
	-	Lina Ratiba Nih	Study the use of hydrogel scaffolds for brain repair after stroke.	(7
	Erythropoi- etin	Yuanfei Wang	Using hyaluronan/methyl cellulose (HAMC) hydrogels for erythropoietin delivery to the brain in order to endogenous stem cell stimulation after stroke.	(4
	Paclitaxel	Sudhir Hulikal Ranga- nath	Fabrication of paclitaxel-loaded PLGA microspheres in or- der to achieve implantable sustained release glioma chemo- therapy.	(7
	Cyclosporine A Curcumin	Matthew J. Caicco Xi Chen	Using parenteral PLGA/HAMC composite for sustained brain delivery of cyclosporine A to treat stroke. Preparation of a Poloxamer 188-based thermosensitive hy-	
Tissue engineer ing		Kuen Yong Lee	drogel to enhance brain targeted delivery of curcumin. Guiding tissue formation in mechanically stressed environ- ments by hydrogels.	(8
•	Different tis- sues	Jeanie L. Drury	Guiding tissue formation in mechanically stressed environ- ments by hydrogels.	(8

.....

Route of Adminis- tration	Loaded Drug	First Author	The major goal of the study	Ref.
	Cartilage tissue	Nguyen, Kytai Truong	The use of photopolymerizable hydrogels in tissue engineer- ing.	(84)
	Vascular tissue	Ali Khademhosseini	The use of microengineered hydrogels for tissue engineering.	(44)
	Soft tissues	Junmin Zhu	The use of bioactive polyethylene glycol hydrogels as tissue engineering scaffolds.	(85)
	Damaged tis- sue	S. Sayyar	The use of chitosan/graphene composite hydrogels for tissue engineering	(86)
	adipose, bone, cartilage, intervertebral discs and muscle tissue	A. Sivashanmugan	The application of injectable hydrogels in tissue engineering.	(43)
	Vascular, muscular, and neural tissues	Jeroen Leijten	Study of the spatiotemporal control over biomaterials for tis- sue engineering purposes.	(87)
	Dermal, heart valve, vascu- lar, corneal and esopha- geal tissue	Sheva Naahidi	Design of non-invasive, biocompatible, and smart hydrogels for tissue engineering purposes.	(88)
Gene delivery	GFP express- ing plasmid	Mariam Mohammadi	Synthesis of folic acid coupled poly(L-lactide)-b- poly(ethylene glycol) and three-layered micelles for gene delivery to activate macrophages in rheumatoid arthritis treatment.	(89)
	Vascular endothe- lial growth factor-165 (VEGF) pro- angiogenic gene	Arghya Paul	Preparation of injectable and biocompatible methacrylated gelatin hydrogel to deliver polyethylenimine functionalized graphene oxide nanosheets complexed with DNAVEGF to promote vacuologenesis and cardiac repair for myocardial therapy.	(90)
	Anti-onco- gene	Yi Yang	Preparation of a biodegradable folate-poly(ester amine) polymer and a PECE thermosensitive hydrogel composite for sustained gene delivery to increase anti-tumor effect.	(91)
	Non-viral vectors	Jeremy Zhang	Using poloxamin/fibrin hybrid hydrogels to induce localized and controlled non-viral gene delivery.	(92)
	Calcium phosphate- DNA	Melissa D. Krebs	Using injectable alginate hydrogels for gene delivery of cal- cium phosphate DNA nanoparticles in order to promote os- teogenesis.	(93)

Cor	Continued Table 1.					
	Route of	Loaded		First Author	The major goal of the study	Ref.
	Adminis- tration	Drug				
		VEGF	and	Yuguo Lei	Synthesis of poly ethylene imine /DNA polyplexes (by caged	(45)
		βgal			nanoparticle encapsulation (CnE) technique) into hyaluronic	
					acid and fibrin hydrogels for local gene delivery purposes.	

of insulin to overcome the complications associated with parenteral insulin administration (36).

### 6.2. Parenteral route

For many drugs such as peptide and protein, parenteral route is the most favorite route of administration. Hydrogels as controlled drug delivery systems could be used for parenteral drug delivery. Hydrogels can be used to prolong and sustain drug release, increase drug half-life, increase bioavailability, drug protection against enzymatic degradation, decrease frequency of drug administration and so increase patient compliance. Injectable hydrogels are usually temperature sensitive so they are sol (fluid) at room temperature and gel (viscous) at body temperature. The gelation process is able to sustain drug release and improve drug bioavailability. Poloxamer-based hydrogels are the most common temperature sensitive hydrogels which were used in parenteral drug delivery systems, but their limitation is the lack of biodegradability (37).

### 6.3. Nasal route

Nasal drug delivery has the advantages of high patient compliance and prevention of hepatic first pass effect which could increase drug bioavailability. However, this route of delivery also has its own limitation including barrier action to the absorption of the macromolecules across the mucosal membranes and short nasal residence time because of mucosal turnover. Hydrogels with mucoadhesive, viscoelastic and biocompatible properties (such as chitosan hydrogels) are highly considered as novel carriers for nasal delivery which could increase nasal residence time of loaded active ingredients. Most of the polymers used in nasal delivery systems are thermosensitive and capable to form gel at site of action at body temperature (38).

# 6.4. Ocular route

Ocular drug delivery via eye drops is a common route of topical administration which is not efficient enough and may cause systemic adverse reactions. Only about 5 % of the incorporated drug could reach to the intraocular tissue, but about 95 % would be lost through tear drainage. Also, the residence time of drug in the eye is very short. So, the development of novel drug delivery systems to increase drug residence time and decrease drug loss and systemic adverse effect, would be highly desirable (39). Many researchers considered contact lenses as ocular drug delivery systems which have the advantage of increasing drug residence time and drug bioavailability. Hydrogel contact lenses because of their transparency and biocompatibility were widely considered as ocular drug delivery systems. In this regard drug molecules could be homogenously dispersed in hydrogel matrices (such as HEMA polymers), but this method is only limited to hydrophilic drugs and may cause fast drug release (39). In order to solve this problem, hydrophobic or ionic monomers could be incorporated to HEMA hydrogels to increase drug-hydrogel interaction and control drug release rate and also could increase drug loading capacity (40).

### 6.5. Topical route

Topical drug delivery is one of the favorite routes of administration which is used to reduce adverse effects and to localize high amounts of drug at target site. Hydrogels because of the low toxicity potential and sustained drug release are considered as suitable carriers for topical drug delivery. Also, hydrogels have the advantage of biocompatibility, softness and high water content which could mimic natural tissues properties and because of their swelling and hydrating capability, could avoid irritation to enclosed tissues. The other important advantage of hydrogels is the ability of drug protection against harsh environmental conditions (41).

# 6.6. Brain delivery

Drug delivery to brain is still associated with so many challenges and the most important problem is the presence of blood brain barrier. Using implants within the brain could be used for local drug delivery, but have the disadvantage of brain tissue damage and infection. Another approach for brain local delivery is epi-cortical delivery using hydrogels which could release the loaded drug directly to the brain with negligible tissue damage (42).

### 6.7. Tissue engineering

Hydrogels have many advantages for tissue engineering purposes such as similarity to the extracellular matrices of tissues, induction of cell proliferation, negligible irritation to adjacent tissues and sustained release of incorporated growth factors. Injectable hydrogels have superiorities in comparison to other conventional scaffolds (preformed hydrogels) like the ease of handling, deeper penetration to tissues, better margin adaptation and less invasiveness (43). Micro-engineered hydrogels are other promising engineering tools to overcome current challenges facing tissue engineering (44).

### 6.8. Gene delivery

Gene delivery through hydrogel scaffold is delivery of DNA or RNA for the purpose of genetic modification. Hydrogels are capable to increase gene therapy efficacy especially in cancer therapy. In cancer therapy, siRNA or the lethal genes would be encapsulated within hydrogel scaffolds and promote apoptosis of cancerous cells. Gene delivery through hydrogels also have some limitations such as limited gene loading capacity and rapid expulsion of encapsulated genes. Several methods have been considered to overcome these limitations like the condensation of DNA or RNA in nanoparticulate systems and then encapsulate them in a hydrogel scaffold (45).

# 7. Hydrogel limitations and possible approaches for each limitation

During this study numerous papers dealing with hydrogel based drug delivery systems were reviewed to address different limitations of hydrogels. Various approaches for each group of these limitations are summarized in Table 2.

### 8. Conclusion

Although hydrogel with different chemical

Table 2. Most common limitations of hydrogels as drug delivery systems and possible approaches.

	Limitations	Solutions
1	Non-biocompatible and non-biodegradable properties of some hydrogels	• Development of biocompatible and biodegradable hydrogels in- cluding PEG-PLGA-PEGa or using polymers that have hydrolys- able moieties (chemical modification) (94).
2	Too slow responsiveness of stimuli-sensitive hydro- gels	• Making thinner and smaller hydrogels which are fast-acting (3).
3	A rapid burst drug release during hydrogel swelling and fast drug release from large porous hydrogels	• Covalently or physically linking of the drug to the polymer chains prior to gelation (tethering method) (95).
		• Preparation of di-block or tri-block copolymers by covalent cross linking method to overcome rapid dissolution and fast drug release (96).
4	Possibility of drug deactivation and initial burst re- lease in entrapment method	• Application of tethering method (95).
5	Possibility of drug deactivation during polymer bind- ing in covalent binding method	• Using appropriate linkers, in which drug release could be tuned, instead of direct covalent binding of drug to polymer.

	Limitations	Solutions
6	Non-specific drug release mechanism in diffusion- controlled release hydrogels	• Using chemically and biologically stimulated release triggers to control the drug release from hydrogels (95).
7	Potential toxicity of residual unreacted small-mol- ecule cross-linkers in small molecule cross-linking method for hydrogel making	• Using polymer-polymer cross-linking method by formation of Schiff base or Michael addition reaction (96).
8		• Introducing hydrophobic domains directly into the hydrogenetwork.
		• Formation of a solid molecular dispersion of poorly soluble drug.
		• Encapsulation of drug in polymer nanoparticles to obtain well dispersed drug slurry.
		• Preparation of a novel nanoparticle and hydrogel composited rug delivery system (96, 97).
		• Preparation of lipid-based nanoparticles (solid lipid nanoparticle and nanostructured lipid carriers) as suitable nanocarriers for hy
9	Difficulty of incorporation of drug-loaded colloidal	<ul><li>drophobic drugs (98-100) and then incorporate them in hydrogel.</li><li>Preparation of mixed delivery systems such as Liposome-ir</li></ul>
10	carriers in hydrogels Fast dissolution of chitosan-based hydrogel matrices in stomach acidic pH for oral drug delivery systems	<ul><li>hydrogels (101).</li><li>The formation of poly electrolyte complexes, conjugation of cross-linking</li></ul>
		• Nanoparticle synthesis such as chitosan-poly (g-glycolic acid based hydrogels for oral insulin delivery (36).
11	Lack of biodegradability and toxicity potential of Poloxamer-based thermosensitive hydrogels	• Preparation of PEG-based hydrogels which are cross-linked the hydrophobic polyesters such as PLGA and PCLb which are bid degradable and biocompatible (102).
		• Preparation of PEG-PNIPAAmc copolymer which act as a suit able temperature sensitive in-situ forming hydrogel (102).
12	Slow and varying response of light-responsive hydro- gels to stimulus (light) (55)	• Fabrication of cross-linked polymers of 2-hydroxyethyl methac rylate which were functionalized with azobenzene groups (103).
		• Forming an interpenetrating polymer network between poly acrylamide and poly acrylic acid (104)
13	Clogging of needle during injection of pH sensitive and temperature sensitive hydrogels	• Preparation of dual responsive hydrogels (pH/thermo-sensitiv hydrogels) (55).
14	pH related activity and solubility and slow sol-gel transition phase	• Chemical modification to improve solubility profile and bette mucoadhesiveness (38).
15	Rapid drug release and low drug loading	• Incorporation of hydrophobic monomers such as 4- vinylpyr dine or ionic monomers such as N-(3-aminopropyl) methacry amide to pHEMAd hydrogels in order to increase drug loadin capacity and control drug release rate (40).

Limitations	Solutions
plantation of Pre-formed hydrogels or scaffolds using	• Substitution of injectable hydrogel systems in order to over come these scaffold limitations (43).
	Preparation of micro-engineered hydrogels (44).
The lack of specific cell adhesive properties of PEG hydrogels as scaffold for tissue engineering	• Cell adhesive modification of PEG hydrogels using extrace lular matrix proteins (85).
Low DNA or RNA loading capacity in hydrogel net- work and limited capability of transgene expression for gene delivery purposes	<ul> <li>Incorporation of DNA/polymer polyplexes within PEG hydrogel scaffolds (105).</li> <li>Using hyaluronic acid and fibrin hydrogels as scaffolds for DNA/polymer polyplexes (45).</li> </ul>
The lack of efficiency and suitability as carrier for small molecular weight and hydrophobic active phar- maceuticals	• Synthesis of hydrogel carriers which are fabricated from copolymers of methyl methacrylate and acrylic acid as novel oral dru delivery systems for small molecular weight and hydrophobic active pharmaceuticals (106).
	• Composition of agar/alginate beads which have the advantag
	The risk of infection and the challenge of surgical im- plantation of Pre-formed hydrogels or scaffolds using in tissue engineering The lack of specific cell adhesive properties of PEG hydrogels as scaffold for tissue engineering Low DNA or RNA loading capacity in hydrogel net- work and limited capability of transgene expression for gene delivery purposes The lack of efficiency and suitability as carrier for small molecular weight and hydrophobic active phar-

c Polyethylene glycol - Poly(N-isopropylacrylamide)

d Poly 2-Hydroxyethylmethacrylate

moieties are available for drug delivery but based on the final purpose of delivery system and route of administration we need to well characterize hydrogel forming polymer. Development of successful hydrogel based delivery system is possible upon knowledge about physiochemical properties of the hydrogel forming polymers and understanding the influencing factors which control the swelling behaviors, hydrophilicity, biodegradability, biocompatibility and targetability of the selected polymer. Hydrogels as drug delivery systems have many advantages including biocompatibility, low toxicity and good swelling behavior but depending on chemical moieties of the gel forming polymers and route of administration some limitations would ap-

### 9. References

1. Hamidi M, Azadi A, Rafiei P. Hydrogel nanoparticles in drug delivery. *Adv Drug Deliv Rev.* 2008 Dec 14;60(15):1638-49. doi: 10.1016/j. addr.2008.08.002.

\_\_\_\_\_

2. Hoffman AS. Hydrogels for biomedical applications. *Adv Drug Deliv Rev.* 2002 Jan 17;54(1):3-12. pear in delivery of active pharmaceuticals such as slow responsiveness of stimuli-sensitive hydrogels, possibility of rapid burst drug release, possibility of drug reactivation, limited hydrophobic drug delivery, low mechanical strength, etc. which should be overcome through different approaches that are suggested by different researchers.

### **Ethical approval**

This review article does not include any animal or human studies done by any of the authors.

### **Conflict of Interest**

None declared.

3. Qiu Y, Park K. Environment-sensitive hydrogels for drug delivery. *Adv Drug Deliv Rev.* 2001 Dec 31;53(3):321-39.

4. Omidian H, Park K. Swelling agents and devices in oral drug delivery. *J Drug Deliv Sci Technol*. 2008;18(2):83-93.

5. Peppas NA, Merrill EW. Poly (vinyl alcohol) hydrogels: Reinforcement of radiation-cross-

linked networks by crystallization. *J Polym Sci A Polym Chem.* 1976;14(2):441-57.

6. Campoccia D, Doherty P, Radice M, Brun P, Abatangelo G, Williams DF. Semisynthetic resorbable materials from hyaluronan esterification. *Biomaterials*. 1998;19(23):2101-27.

7. Ahmed EM. Hydrogel: Preparation, characterization, and applications: A review. *J Adv Res.* 2015 Mar;6(2):105-21. doi: 10.1016/j. jare.2013.07.006.

8. Lin C-C, Metters AT. Hydrogels in controlled release formulations: network design and mathematical modeling. *Adv Drug Deliv Rev.* 2006 Nov 30;58(12-13):1379-408.

9. Amsden B. Solute diffusion within hydrogels. Mechanisms and models. *Macromolecules*. 1998;31(23):8382-95.

10. Peppas N, Bures P, Leobandung W, Ichikawa H. Hydrogels in pharmaceutical formulations. *Eur J Pharm Biopharm*. 2000;50(1):27-46.

11. Lin C-C, Anseth KS. PEG hydrogels for the controlled release of biomolecules in regenerative medicine. *Pharm Res.* 2009 Mar;26(3):631-43. doi: 10.1007/s11095-008-9801-2.

12. Bettini R, Colombo P, Massimo G, Catellani PL, Vitali T. Swelling and drug release in hydrogel matrices: polymer viscosity and matrix porosity effects. *Eur J Pharm Sci.* 1994;2(3):213-9.

13. Siepmann J, Peppas N. Modeling of drug release from delivery systems based on hydroxy-propyl methylcellulose (HPMC). *Adv Drug Deliv Rev.* 2012;64:163-74.

14. Ravaine V, Ancla C, Catargi B. Chemically controlled closed-loop insulin delivery. *J Control Release*. 2008 Nov 24;132(1):2-11. doi: 10.1016/j.jconrel.2008.08.009.

15. Das N. Preparation methods and properties of hydrogel: a review. *Int J Pharm Pharm Sci.* 2013;5(3):112-7.

16. Pillai O, Panchagnula R. Polymers in drug delivery. *Curr Opin Chem Biol.* 2001 Aug;5(4):447-51.

17. Esquerdo V, Cadaval T, Dotto G, Pinto L. Chitosan scaffold as an alternative adsorbent for the removal of hazardous food dyes from aqueous solutions. *J Colloid Interface Sci.* 2014 Jun 15;424:7-15. doi: 10.1016/j.jcis.2014.02.028.

18. Kamath KR, Park K. Biodegradable hydrogels in drug delivery. *Adv Drug Deliv Rev.* 

1993;11(1-2):59-84.

19. Bulpitt P, Aeschlimann D. New strategy for chemical modification of hyaluronic acid: preparation of functionalized derivatives and their use in the formation of novel biocompatible hydrogels. *J Biomed Mater Res.* 1999 Nov;47(2):152-69.

20. Santo VE, Frias AM, Carida M, Cancedda R, Gomes ME, Mano JF, et al. Carrageenan-based hydrogels for the controlled delivery of PDGF-BB in bone tissue engineering applications. *Biomacromolecules*. 2009;10(6):1392-401.

21. YOTSUYANAGI T, OHKUBO T, OHHASHI T, Ikeda K. Calcium-induced gelation of alginic acid and pH-sensitive reswelling of dried gels. *Chem Pharm Bull.* 1987;35(4):1555-63.

22. Teles H, Vermonden T, Eggink G, Hennink W, de Wolf F. Hydrogels of collagen-inspired telechelic triblock copolymers for the sustained release of proteins. *J Control Release*. 2010 Oct 15;147(2):298-303. doi: 10.1016/j.jconrel.2010.07.098.

23. Rao JK, Ramesh DV, Rao KP. Implantable controlled delivery systems for proteins based on collagen—pHEMA hydrogels. *Biomaterials*. 1994 Apr;15(5):383-9.

24. Almany L, Seliktar D. Biosynthetic hydrogel scaffolds made from fibrinogen and polyethylene glycol for 3D cell cultures. *Biomaterials*. 2005;26(15):2467-77.

25. Metters A, Anseth K, Bowman C. Fundamental studies of a novel, biodegradable PEG-*b*-PLA hydrogel. *Polymer*. 2000;41(11):3993-4004.

26. Mohammadi-Samani S, Taghipour B. PLGA micro and nanoparticles in delivery of peptides and proteins; problems and approaches. *Pharm Dev Technol.* 2015 Jun;20(4):385-93. doi: 10.3109/10837450.2014.882940.

27. Jiang S, Liu S, Feng W. PVA hydrogel properties for biomedical application. *J Mech Behav Biomed Mater*. 2011 Oct;4(7):1228-33. doi: 10.1016/j.jmbbm.2011.04.005.

28. Li X, Li J. Supramolecular hydrogels based on inclusion complexation between poly (ethylene oxide)-b-poly ( $\varepsilon$ -caprolactone) diblock copolymer and  $\alpha$ -cyclodextrin and their controlled release property. *J Biomed Mater Res A*. 2008 Sep 15;86(4):1055-61.

29. Dash TK, Konkimalla VB. Poly- $\epsilon$ caprolactone based formulations for drug delivery and tissue engineering: A review. *J Control Re*- *lease.* 2012 Feb 28;158(1):15-33. doi: 10.1016/j. jconrel.2011.09.064.

30. Reza AT, Nicoll SB. Characterization of novel photocrosslinked carboxymethylcellulose hydrogels for encapsulation of nucleus pulposus cells. *Acta Biomater*. 2010 Jan;6(1):179-86. doi: 10.1016/j.actbio.2009.06.004.

31. Wang W, Wang J, Kang Y, Wang A. Synthesis, swelling and responsive properties of a new composite hydrogel based on hydroxyethyl cellulose and medicinal stone. *Compos Part B-Eng.* 2011;42(4):809-18.

32. Sannino A, Demitri C, Madaghiele M. Biodegradable cellulose-based hydrogels: design and applications. *Materials*. 2009;2(2):353-73.

33. Lott JR, McAllister JW, Arvidson SA, Bates FS, Lodge TP. Fibrillar structure of methylcellulose hydrogels. *Biomacromolecules*. 2013 Aug 12;14(8):2484-8. doi: 10.1021/bm400694r.

34. Liu X, Zhou Y, Nie W, Song L, Chen P. Fabrication of hydrogel of hydroxypropyl cellulose (HPC) composited with graphene oxide and its application for methylene blue removal. *J Mater Sci.* 2015;50(18):6113-23.

35. Morishita M, Goto T, Nakamura K, Lowman AM, Takayama K, Peppas NA. Novel oral insulin delivery systems based on complexation polymer hydrogels: Single and multiple administration studies in type 1 and 2 diabetic rats. *J Control Release*. 2006 Feb 21;110(3):587-94.

36. Chaturvedi K, Ganguly K, Nadagouda MN, Aminabhavi TM. Polymeric hydrogels for oral insulin delivery. *J Control Release*. 2013 Jan 28;165(2):129-38. doi: 10.1016/j.jcon-rel.2012.11.005.

37. Alexander A, Khan J, Saraf S, Saraf S. Polyethylene glycol (PEG)–Poly (N-isopropylacrylamide)(PNIPAAm) based thermosensitive injectable hydrogels for biomedical applications. *Eur J Pharm Biopharm.* 2014 Nov;88(3):575-85. doi: 10.1016/j.ejpb.2014.07.005

38. Nazar H, Fatouros DG, van der Merwe SM, Bouropoulos N, Avgouropoulos G, Tsibouklis J, et al. Thermosensitive hydrogels for nasal drug delivery: The formulation and characterisation of systems based on N-trimethyl chitosan chloride. *Eur J Pharm Biopharm.* 2011 Feb;77(2):225-32. doi: 10.1016/j.ejpb.2010.11.022.

39. Gulsen D, Chauhan A. Dispersion of microemulsion drops in HEMA hydrogel: a potential

ophthalmic drug delivery vehicle. *Int J Pharm.* 2005 Mar 23;292(1-2):95-117.

40. Hu X, Hao L, Wang H, Yang X, Zhang G, Wang G, et al. Hydrogel contact lens for extended delivery of ophthalmic drugs. *Int J Polym Sci.* 2011;2011.

41. Morsi NM, Abdelbary GA, Ahmed MA. Silver sulfadiazine based cubosome hydrogels for topical treatment of burns: development and in vitro/in vivo characterization. *Eur J Pharm Biopharm*. 2014 Feb;86(2):178-89. doi: 10.1016/j. ejpb.2013.04.018.

42. Wang Y, Cooke MJ, Morshead CM, Shoichet MS. Hydrogel delivery of erythropoietin to the brain for endogenous stem cell stimulation after stroke injury. *Biomaterials*. 2012;33(9):2681-92. doi: 10.1016/j.biomaterials.2011.12.031.

43. Sivashanmugam A, Kumar RA, Priya MV, Nair SV, Jayakumar R. An overview of injectable polymeric hydrogels for tissue engineering. *Eur Polym J.* 2015;72:543-65.

44. Khademhosseini A, Langer R. Microengineered hydrogels for tissue engineering. *Biomaterials*. 2007;28(34):5087-92.

45. Lei Y, Rahim M, Ng Q, Segura T. Hyaluronic acid and fibrin hydrogels with concentrated DNA/PEI polyplexes for local gene delivery. *J Control Release*. 2011 Aug 10;153(3):255-61. doi: 10.1016/j.jconrel.2011.01.028.

46. Colombo P. Swelling-controlled release in hydrogel matrices for oral route. *Adv Drug Deliv Rev.* 1993;11(1-2):37-57.

47. Knuth K, Amiji M, Robinson JR. Hydrogel delivery systems for vaginal and oral applications: Formulation and biological considerations. *Adv Drug Deliv Rev.* 1993;11(1-2):137-67.

48. Shalaby WS, Blevins WE, Park K. In vitro and in vivo studies of enzyme-digestible hydrogels for oral drug delivery. *J Control Release*. 1992;19(1-3):131-44.

49. Kim B, Peppas NA. In vitro release behavior and stability of insulin in complexation hydrogels as oral drug delivery carriers. *Int J Pharm*. 2003 Nov 6;266(1-2):29-37.

50. Şenel S, Ikinci G, Kaş S, Yousefi-Rad A, Sargon M, Hıncal A. Chitosan films and hydrogels of chlorhexidine gluconate for oral mucosal delivery. *Int J Pharm.* 2000 Jan 5;193(2):197-203.

51. Kim KS, Park SJ, Yang JA, Jeon JH, Bhang SH, Kim BS, et al. Injectable hyaluronic acid-ty-

ramine hydrogels for the treatment of rheumatoid arthritis. *Acta Biomater*. 2011 Feb;7(2):666-74. doi: 10.1016/j.actbio.2010.09.030.

52. Purcell BP, Lobb D, Charati MB, Dorsey SM, Wade RJ, Zellars KN, et al. Injectable and bioresponsive hydrogels for on-demand matrix metalloproteinase inhibition. *Nat Mater*. 2014 Jun;13(6):653-61. doi: 10.1038/nmat3922.

53. Bisharat L, Perinelli DR, Berardi A, Bonacucina G, Logrippo S, Elhajji FWD, et al. Influence of Testing Parameters on In Vitro Tramadol Release from Poloxamer Thermogels using the Immersion Cell Method. *AAPS PharmSciTech*. 2017 Oct;18(7):2706-2716. doi: 10.1208/s12249-017-0753-x.

54. Jensen SS, Jensen H, Møller EH, Cornett C, Siepmann F, Siepmann J, et al. In vitro release studies of insulin from lipid implants in solution and in a hydrogel matrix mimicking the subcutis. *Eur J Pharm Sci.* 2016 Jan 1;81:103-12. doi: 10.1016/j.ejps.2015.10.011.

55. Oak M, Mandke R, Singh J. Smart polymers for peptide and protein parenteral sustained delivery. *Drug Discov Today Technol*. 2012 Summer;9(2):e71-e174. doi: 10.1016/j. ddtec.2012.05.001.

56. Vermonden T, Censi R, Hennink WE. Hydrogels for protein delivery. *Chem Rev.* 2012 May 9;112(5):2853-88. doi: 10.1021/cr200157d.

57. Wu J, Wei W, Wang LY, Su ZG, Ma GH. A thermosensitive hydrogel based on quaternized chitosan and poly(ethylene glycol) for nasal drug delivery system. *Biomaterials*. 2007 Apr;28(13):2220-32.

58. Luppi B, Bigucci F, Cerchiara T, Zecchi V. Chitosan-based hydrogels for nasal drug delivery: from inserts to nanoparticles. *Expert Opin Drug Deliv.* 2010 Jul;7(7):811-28. doi: 10.1517/17425247.2010.495981.

59. Xu X, Shen Y, Wang W, Sun C, Li C, Xiong Y, et al. Preparation and in vitro characterization of thermosensitive and mucoadhesive hydrogels for nasal delivery of phenylephrine hydrochloride. *Eur J Pharm Biopharm.* 2014 Nov;88(3):998-1004. doi: 10.1016/j.ejpb.2014.08.015.

60. Alsarra IA, Hamed AY, Alanazi FK, Neau SH. Rheological and mucoadhesive characterization of poly (vinylpyrrolidone) hydrogels designed for nasal mucosal drug delivery. *Arch Pharm Res.* 2011 Apr;34(4):573-82. doi: 10.1007/s12272-011-

0407-6.

61. Alsarra IA, Hamed AY, Mahrous GM, El Maghraby GM, Al-Robayan AA, Alanazi FK. Mucoadhesive polymeric hydrogels for nasal delivery of acyclovir. *Drug Dev Ind Pharm.* 2009 Mar;35(3):352-62. doi: 10.1080/03639040802360510.

62. Luppi B, Bigucci F, Abruzzo A, Corace G, Cerchiara T, Zecchi V. Freeze-dried chitosan/pectin nasal inserts for antipsychotic drug delivery. *Eur J Pharm Biopharm.* 2010 Aug;75(3):381-7. doi: 10.1016/j.ejpb.2010.04.013.

63. Zignani M, Tabatabay C, Gurny R. Topical semi-solid drug delivery: kinetics and tolerance of ophthalmic hydrogels. *Adv Drug Deliv Rev.* 1995;16(1):51-60.

64. Ribeiro A, Veiga F, Santos D, Torres-Labandeira JJ, Concheiro A, Alvarez-Lorenzo C. Bioinspired imprinted PHEMA-hydrogels for ocular delivery of carbonic anhydrase inhibitor drugs. *Biomacromolecules*. 2011;12(3):701-9.

65. Xinming L, Yingde C, Lloyd AW, Mikhalovsky SV, Sandeman SR, Howel CA, et al. Polymeric hydrogels for novel contact lens-based ophthalmic drug delivery systems: A review. *Cont Lens Anterior Eye*. 2008 Apr;31(2):57-64.

66. Barbu E, Verestiuc L, Iancu M, Jatariu A, Lungu A, Tsibouklis J. Hybrid polymeric hydrogels for ocular drug delivery: nanoparticulate systems from copolymers of acrylic acid-functionalized chitosan and N-isopropylacrylamide or 2-hydroxyethyl methacrylate. *Nanotechnology.* 2009 Jun 3;20(22):225108. doi: 10.1088/0957-4484/20/22/225108.

67. Yin H, Gong C, Shi S, Liu X, Wei Y, Qian Z. Toxicity evaluation of biodegradable and thermosensitive PEG-PCL-PEG hydrogel as a potential in situ sustained ophthalmic drug delivery system. *J Biomed Mater Res B Appl Biomater.* 2010 Jan;92(1):129-37. doi: 10.1002/jbm.b.31498.

68. Şenyiğit T, Tekmen I, Sönmez Ü, Santi P, ÖzerÖ. Deoxycholate hydrogels of betamethasone-17-valerate intended for topical use: In vitro and in vivo evaluation. *Int J Pharm*. 2011 Jan 17;403(1-2):123-9. doi: 10.1016/j.ijpharm.2010.10.036.

69. Valenta C, Nowack E, Bernkop-Schnürch A. Deoxycholate-hydrogels: novel drug carrier systems for topical use. *Int J Pharm.* 1999 Aug 5;185(1):103-11.

70. Cerchiara T, Luppi B, Bigucci F, Orienti

I, Zecchi V. Physically cross-linked chitosan hydrogels as topical vehicles for hydrophilic drugs. *J Pharm Pharmacol.* 2002 Nov;54(11):1453-9.

71. Sanna V, Gavini E, Cossu M, Rassu G, Giunchedi P. Solid lipid nanoparticles (SLN) as carriers for the topical delivery of econazole nitrate: invitro characterization, ex-vivo and in-vivo studies. *J Pharm Pharmacol.* 2007 Aug;59(8):1057-64.

72. Fang J-Y, Leu Y-L, Wang Y-Y, Tsai Y-H. In vitro topical application and in vivo pharmacodynamic evaluation of nonivamide hydrogels using Wistar rat as an animal model. *Eur J Pharm Sci.* 2002 Jun;15(5):417-23.

73. Yoncheva K, Doytchinova I, Tankova L. Preparation and evaluation of isosorbide mononitrate hydrogels for topical fissure treatment. *Curr Drug Deliv.* 2012 Sep;9(5):452-8.

74. Murphy DJ, Sankalia MG, Loughlin RG, Donnelly RF, Jenkins MG, McCarron PA. Physical characterisation and component release of poly (vinyl alcohol)–tetrahydroxyborate hydrogels and their applicability as potential topical drug delivery systems. *Int J Pharm.* 2012 Feb 28;423(2):326-34. doi: 10.1016/j.ijpharm.2011.11.018.

75. Özcan İ, Abacı Ö, Uztan AH, Aksu B, Boyacıoğlu H, Güneri T, et al. Enhanced topical delivery of terbinafine hydrochloride with chitosan hydrogels. *AAPS PharmSciTech*. 2009;10(3):1024-31. doi: 10.1208/s12249-009-9299-x.

76. Plant GW, Woerly S, Harvey AR. Hydrogels containing peptide or aminosugar sequences implanted into the rat brain: influence on cellular migration and axonal growth. *Exp Neurol*. 1997 Feb;143(2):287-99.

77. Bjugstad K, Lampe K, Kern D, Mahoney M. Biocompatibility of poly (ethylene glycol)based hydrogels in the brain: An analysis of the glial response across space and time. *J Biomed Mater Res A*. 2010 Oct;95(1):79-91. doi: 10.1002/ jbm.a.32809.

78. Nih LR, Carmichael ST, Segura T. Hydrogels for brain repair after stroke: an emerging treatment option. *Curr Opin Biotechnol.* 2016 Aug;40:155-163. doi: 10.1016/j. copbio.2016.04.021.

79. Ranganath SH, Kee I, Krantz WB, Chow PK-H, Wang C-H. Hydrogel matrix entrapping PLGA-paclitaxel microspheres: drug delivery with near zero-order release and implantability advantages for malignant brain tumour chemo-

therapy. *Pharm Res.* 2009 Sep;26(9):2101-14. doi: 10.1007/s11095-009-9922-2.

80. Caicco MJ, Cooke MJ, Wang Y, Tuladhar A, Morshead CM, Shoichet MS. A hydrogel composite system for sustained epi-cortical delivery of Cyclosporin A to the brain for treatment of stroke. *J Control Release*. 2013 Mar 28;166(3):197-202. doi: 10.1016/j.jconrel.2013.01.002.

81. Chen X, Zhi F, Jia X, Zhang X, Ambardekar R, Meng Z, et al. Enhanced brain targeting of curcumin by intranasal administration of a thermosensitive poloxamer hydrogel. *J Pharm Pharmacol.* 2013 Jun;65(6):807-16. doi: 10.1111/ jphp.12043.

82. Lee KY, Mooney DJ. Hydrogels for tissue engineering. *Chem Rev.* 2001 Jul;101(7):1869-79.

83. Drury JL, Mooney DJ. Hydrogels for tissue engineering: scaffold design variables and applications. *Biomaterials*. 2003;24(24):4337-51.

84. Nguyen KT, West JL. Photopolymerizable hydrogels for tissue engineering applications. *Biomaterials*. 2002 Nov;23(22):4307-14.

85. Zhu J. Bioactive modification of poly (ethylene glycol) hydrogels for tissue engineering. *Biomaterials*. 2010 Jun;31(17):4639-56. doi: 10.1016/j.biomaterials.2010.02.044.

86. Sayyar S, Murray E, Thompson B, Chung J, Officer DL, Gambhir S, et al. Processable conducting graphene/chitosan hydrogels for tissue engineering. *J Mater Chem B*. 2015;3(3):481-90.

87. Leijten J, Seo J, Yue K, Trujillo-de Santiago G, Tamayol A, Ruiz-Esparza GU, et al. Spatially and temporally controlled hydrogels for tissue engineering. *Mater Sci Eng R Rep.* 2017 Sep;119:1-35. doi: 10.1016/j.mser.2017.07.001.

88. Naahidi S, Jafari M, Logan M, Wang Y, Yuan Y, Bae H, et al. Biocompatibility of hydrogel-based scaffolds for tissue engineering applications. *Biotechnol Adv.* 2017 Sep;35(5):530-544. doi: 10.1016/j.biotechadv.2017.05.006.

89. Mohammadi M, Li Y, Abebe DG, Xie Y, Kandil R, Kraus T, et al. Folate receptor targeted three-layered micelles and hydrogels for gene delivery to activated macrophages. *J Control Release*. 2016 Dec 28;244(Pt B):269-279. doi: 10.1016/j.jconrel.2016.08.020.

90. Paul A, Hasan A, Kindi HA, Gaharwar AK, Rao VT, Nikkhah M, et al. Injectable graphene oxide/hydrogel-based angiogenic gene delivery system for vasculogenesis and cardiac re-

pair. ACS Nano. 2014 Aug 26;8(8):8050-62. doi: 10.1021/nn5020787.

91. Yang Y, Zhao H, Jia Y, Guo Q, Qu Y, Su J, et al. A novel gene delivery composite system based on biodegradable folate-poly (ester amine) polymer and thermosensitive hydrogel for sustained gene release. *Sci Rep.* 2016 Feb 17;6:21402. doi: 10.1038/srep21402.

92. Zhang J, Sen A, Cho E, Lee JS, Webb K. Poloxamine/fibrin hybrid hydrogels for non-viral gene delivery. *J Tissue Eng Regen Med.* 2017 Jan;11(1):246-255. doi: 10.1002/term.1906.

93. Krebs MD, Salter E, Chen E, Sutter KA, Alsberg E. Calcium phosphate-DNA nanoparticle gene delivery from alginate hydrogels induces in vivo osteogenesis. *J Biomed Mater Res A*. 2010 Mar 1;92(3):1131-8. doi: 10.1002/jbm.a.32441.

94. Li Z, Ning W, Wang J, Choi A, Lee P-Y, Tyagi P, et al. Controlled gene delivery system based on thermosensitive biodegradable hydrogel. *Pharm Res.* 2003 Jun;20(6):884-8.

95. Bhattarai N, Gunn J, Zhang M. Chitosanbased hydrogels for controlled, localized drug delivery. *Adv Drug Deliv Rev.* 2010 Jan 31;62(1):83-99. doi: 10.1016/j.addr.2009.07.019.

96. Hoare TR, Kohane DS. Hydrogels in drug delivery: progress and challenges. *Polymer*. 2008;49(8):1993-2007.

97. Gou M, Li X, Dai M, Gong C, Wang X, Xie Y, et al. A novel injectable local hydrophobic drug delivery system: Biodegradable nanoparticles in thermo-sensitive hydrogel. *Int J Pharm.* 2008 Jul 9;359(1-2):228-33. doi: 10.1016/j. ijpharm.2008.03.023.

98. Ghasemiyeh P, Azadi A, Daneshamouz S, Heidari R, Azarpira N, Mohammadi-Samani S. Cyproterone Acetate-Loaded Nanostructured Lipid Carriers: Effect of Particle Size on Skin Penetration and Follicular Targeting. *Pharm Dev Technol.* 2019 Mar 19:1-12. doi: 10.1080/10837450.2019.1596133.

99. Ghasemiyeh P, Azadi A, Daneshamouz S, Samani SM. Cyproterone acetate-loaded solid lipid nanoparticles (SLNs) and nanostructured lipid carriers (NLCs): preparation and optimization. *Trends* 

# in Pharmaceutical Sciences. 2017;3(4):275-86.

100. Ghasemiyeh P, Mohammadi-Samani S. Solid lipid nanoparticles and nanostructured lipid carriers as novel drug delivery systems: applications, advantages and disadvantages. *Res Pharm Sci.* 2018 Aug;13(4):288-303. doi: 10.4103/1735-5362.235156.

101. Hurler J, Berg OA, Skar M, Conradi AH, Johnsen PJ, Skalko-Basnet N. Improved burns therapy: liposomes-in-hydrogel delivery system for mupirocin. *J Pharm Sci.* 2012 Oct;101(10):3906-15. doi: 10.1002/jps.23260.

102. Alexander A, Khan J, Saraf S, Saraf S. Poly (ethylene glycol)–poly (lactic-co-glycolic acid) based thermosensitive injectable hydrogels for biomedical applications. *J Control Release*. 2013 Dec 28;172(3):715-29. doi: 10.1016/j.jconrel.2013.10.006.

103. Unger K, Salzmann P, Masciullo C, Cecchini M, Koller G, Coclite AM. Novel Light-Responsive Biocompatible Hydrogels Produced by Initiated Chemical Vapor Deposition. *ACS Appl Mater Interfaces*. 2017 May 24;9(20):17408-17416. doi: 10.1021/acsami.7b01527.

104. Xu Y, Ghag O, Reimann M, Sitterle P, Chatterjee P, Nofen E, et al. Development of visible-light responsive and mechanically enhanced "smart" UCST interpenetrating network hydrogels. *Soft Matter*. 2017 Dec 20;14(1):151-160. doi: 10.1039/c7sm01851g.

105. Lei Y, Huang S, Sharif-Kashani P, Chen Y, Kavehpour P, Segura T. Incorporation of active DNA/cationic polymer polyplexes into hydrogel scaffolds. *Biomaterials*. 2010 Dec;31(34):9106-16. doi: 10.1016/j.biomaterials.2010.08.016.

106. Caldorera-Moore M, Maass K, Hegab R, Fletcher G, Peppas N. Hybrid responsive hydrogel carriers for oral delivery of low molecular weight therapeutic agents. *J Drug Deliv Sci Technol.* 2015 Dec 1;30(Pt B):352-359.

107. Yin Z-C, Wang Y-L, Wang K. A pH-responsive composite hydrogel beads based on agar and alginate for oral drug delivery. *J Drug Deliv Sci Technol.* 2018;43:12-8.